Progressive familial intrahepatic cholestasis
Emmanuel Jacquemin

Summary
Progressive familial intrahepatic cholestasis (PFIC) refers to a heterogeneous group of autosomal-recessive disorders of childhood that disrupt bile formation and present with cholestasis of hepatocellular origin. The exact prevalence remains unknown, but the estimated incidence varies between 1/50,000 and 1/100,000 births. Three types of PFIC have been identified and associated with mutations in hepatocellular transport-system genes involved in bile formation. PFIC1 and PFIC2 usually appear in the first months of life, whereas onset of PFIC3 may arise later in infancy, in childhood or even during young adulthood. The main clinical manifestations include cholestasis, pruritus and jaundice. PFIC patients usually develop fibrosis and end-stage liver disease before adulthood. Serum gamma-glutamyltransferase (GGT) activity is normal in PFIC1 and PFIC2 patients, but is elevated in PFIC3 patients. Both PFIC1 and PFIC2 are caused by impaired bile salt secretion due to defects in \(\text{ATP8B1}\) encoding the FIC1 protein and in \(\text{ABCB11}\) encoding the bile salt export pump (BSEP) protein, respectively. Defects in \(\text{ABCB4}\), encoding multidrug resistance 3 protein (MDR3), impair biliary phospholipid secretion, resulting in PFIC3. Diagnosis is based on clinical manifestations, liver ultrasonography, cholangiography and liver histology, as well as on specific tests to exclude other causes of childhood cholestasis. MDR3 and BSEP liver immunostaining, and analysis of biliary lipid composition should help to select PFIC candidates for whom genotyping could be proposed to confirm the diagnosis. Antenatal diagnosis may be proposed for affected families in which a mutation has been identified. Ursodeoxycholic acid (UDCA) therapy should be initiated in all patients to prevent liver damage. In some PFIC1 and PFIC2 patients, biliary diversion may also relieve pruritus and slow disease progression. However, most PFIC patients are ultimately candidates for liver transplantation. Monitoring of liver tumors, especially in PFIC2 patients, should be offered from the first year of life. Hepatocyte transplantation, gene therapy and specific targeted pharmacotherapy may represent alternative treatments in the future.

© 2012 Elsevier Masson SAS. All rights reserved.
Definition
Progressive familial intrahepatic cholestasis (PFIC) refers to a heterogeneous group of autosomal-recessive liver disorders of childhood in which cholestasis of hepatocellular origin often precedes in the neonatal period or first year of life and leads to death from liver failure at ages usually ranging from infancy to adolescence [1,2]. Recent molecular and genetic studies have allowed the identification of genes responsible for three types of PFIC: progressive familial intrahepatic cholestasis type 1 (PFIC1); progressive familial intrahepatic cholestasis type 2 (PFIC2); and progressive familial intrahepatic cholestasis type 3 (PFIC3). It has been shown that all three types are related to mutations in hepatocellular transport-system genes involved in bile formation (Fig. 1).

Epidemiology
In our experience, PFIC represents the cause of cholestasis in 10-15% of children, and 10-15% of liver transplantation indications in children. PFIC1 and PFIC2 represent two-thirds of cases of PFIC, and PFIC3 the remaining third of cases [1]. The true incidence of PFIC is not precisely known, but it is considered a rare disease, with an estimated incidence of 1/50,000 to 1/100,000 births. All types of PFIC are seen worldwide. Both genders appear to be equally affected.

Clinical description
The main characteristics of PFIC types 1-3 are summarized in Table 1. Cholestasis is a major clinical sign in all forms of PFIC. It usually appears in the first months of life in patients with PFIC1 and is characterized by recurrent episodes of jaundice, which become permanent later on in the course of the disease. In cases of PFIC2, the initial presentation and its evolution appear to be more severe, with permanent jaundice from the first months of life and the rapid appearance of liver failure within the first years of life. Early hepatocellular carcinoma (before 1 year of age) and cholangiocarcinoma may complicate the course of PFIC2. Severe pruritus is usually observed in PFIC1 and PFIC2 patients. Phenotypic findings in PFIC1 and PFIC2 are also similar, although slight phenotypic differences have been identified [1-3]. Extrareplicative features, for example, reported in PFIC1 patients (persistent short stature, sensorineural deafness, watery diarrhea, pancreatitis, elevated sweat electrolyte concentration, liver steatosis) have not been described in PFIC2 cases. In contrast to PFIC types 1-2, clinical signs of cholestasis are noted within the first year of life in about one-third of patients and rarely in the neonatal period in PFIC3 patients. PFIC3 may also manifest later in infancy, in childhood and even in young adulthood [4]. Gastrointestinal bleeding due to portal hypertension and cirrhosis are the presenting symptoms in adolescent or young adult patients, and pruritus is usually mild. Progression is characterized by chronic icteric or anicteric cholestasis, portal hypertension and liver failure. In half of all PFIC3 patients, liver transplantation is required at a mean age of 7.5 years. Hepatocarcinoma may also develop during adolescence.

Laboratory findings
Patients with PFIC types 1-2 have normal serum gamma-glutamyltransferase (GGT) activity, normal serum cholesterol levels and very high serum bile acid concentrations. Although PFIC1 and PFIC2 share similar laboratory findings, PFIC2 patients have higher transaminase and alpha-fetoprotein serum levels at diagnosis than those with PFIC1 [1-3]. On the other hand, patients with PFIC3 have a persistent high serum GGT activity, normal serum cholesterol levels and moderately raised concentrations of serum primary bile salts [4].

Histological findings
In PFIC1 patients, liver histology is characterized by canalicular cholestasis and the absence of true ductular proliferation, with only periportal biliary metaplasia of hepatocytes. In PFIC2 patients, the same signs are seen, but the liver architecture is more perturbed, with more pronounced lobular and portal fibrosis and inflammation. Hepatocellular necrosis and giant-cell transformation are also much more pronounced in PFIC2 than in PFIC1, and may persist over time. These differences between PFIC1 and PFIC2 most likely reflect the severe lobular injury present in PFIC2 [3].

In PFIC3, liver histology, obtained at the time of diagnosis, shows portal fibrosis and true ductular proliferation with a mixed inflammatory infiltrate. In a few instances, cholestasis is present in the lobule and in some ductules containing bile plugs. Slight giant-cell transformation of hepatocytes may be observed. Cytokeratin immunostaining confirms the major ductular proliferation within the portal tract. At a later stage, there is extensive portal fibrosis and a typical picture of biliary cirrhosis. Interlobular bile ducts are seen in most portal tracts, and there is neither periductal fibrosis nor biliary epithelium injury [4].
Figure 1  Canalicular transporters involved in progressive familial intrahepatic cholestasis (PFIC) types 1–3: (A) FIC1 maintains the enrichment of aminophospholipids on the inner leaflet of the canalicular membrane, BSEP transports bile acids into bile and MDR3 translocates phosphatidylcholine into bile; (B) PFIC1, PFIC2 and PFIC3 are due to functional defects of FIC1, BSEP and MDR3, respectively. PS: phosphatidylserine; PE: phosphatidylethanolamine; BA: bile acid; PC: phosphatidylcholine.
Etiology

PFIC type 1

PFIC1 (previously reported as “Byler disease”) is caused by mutations in the \( ATP8B1 \) gene (also designated \( FIC1 \)) \cite{1,2,5}. This gene, which encodes a P-type ATPase, is located on human chromosome 18, and is also mutated in the milder phenotype of benign recurrent intrahepatic cholestasis type 1 (BRIC1) and in Greenland familial cholestasis. \( FIC1 \) protein is located on the canalicular membrane of hepatocytes whereas, within the liver, it is mainly expressed in cholangiocytes. This P-type ATPase most likely an aminophospholipid transporter responsible for maintaining the enrichment of phosphatidylserine and phosphatidylethanolamine on the inner leaflet of the plasma membrane. The asymmetrical distribution of lipids in the membrane bilayer plays a protective role against high bile salt concentrations in the canalicular lumen \cite{6}. This flippase may also play a role in maintaining canalicular membrane integrity, including microvilli formation \cite{7,8}. How mutations in this protein cause cholestasis is unclear. It is postulated that abnormal protein function could indirectly disrupt the biliary secretion of bile acids, thereby explaining the low biliary bile acid concentrations found in PFIC1 patients \cite{3}. Different studies have shown that impaired \( ATP8B1 \) function in patients with PFIC1 results in substantial downregulation of the farnesoid X receptor (FXR), a nuclear receptor involved in the regulation of bile acid metabolism, with subsequent downregulation of bile salt export pump (BSEP) protein in the liver, upregulation of bile acid synthesis and apical sodium bile salt transporter (ASBT) in the intestine \cite{9,10}. All in all, these events lead to hepatocyte bile acid overload. Furthermore, evidence of cystic fibrosis transmembrane conductance regulator (CFTR) downregulation in cholangiocytes has also been reported in patients with PFIC1, and could contribute to impairment of bile secretion and explain some of the extrahepatic features \cite{1}.

The \( ATP8B1 \) gene is expressed in various organs, including the liver, pancreas, kidney and small intestine, but is more highly expressed in the small intestine than in the liver. Therefore, it is thought to be involved in the enterohepatic cycling of bile salts. This may also explain the chronic diarrhea present in a few children with PFIC1. Other extrahepatic features associated with PFIC1, such as persistent short stature, deafness and pancreatitis, suggest a general biological cell function for \( FIC1 \) \cite{1,11}.

Table 1 Main characteristics of progressive familial intrahepatic cholestasis (PFIC) types 1–3.

<table>
<thead>
<tr>
<th></th>
<th>PFIC1</th>
<th>PFIC2</th>
<th>PFIC3</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Transmission</strong></td>
<td>Autosomal-recessive</td>
<td>Autosomal-recessive</td>
<td>Autosomal-recessive</td>
</tr>
<tr>
<td><strong>Pruritus</strong></td>
<td>Severe</td>
<td>Very severe</td>
<td>Moderate</td>
</tr>
<tr>
<td><strong>Serum GGT activity</strong></td>
<td>Normal</td>
<td>Normal</td>
<td>High</td>
</tr>
<tr>
<td><strong>Serum ALT</strong></td>
<td>Mildly elevated</td>
<td>&gt;5x normal</td>
<td>Mildly elevated</td>
</tr>
<tr>
<td><strong>Serum AFP</strong></td>
<td>Normal</td>
<td>Elevated</td>
<td>Normal</td>
</tr>
<tr>
<td><strong>Liver histology</strong></td>
<td>Canalicular cholestasis, mild lobular fibrosis</td>
<td>Canalicular cholestasis, lobular fibrosis, giant-cell hepatitis</td>
<td>Ductular proliferation, biliary fibrosis</td>
</tr>
<tr>
<td><strong>Serum primary bile acid concentration</strong></td>
<td>Very high</td>
<td>Very high</td>
<td>High</td>
</tr>
<tr>
<td><strong>Bile composition</strong></td>
<td>Low primary bile acid concentrations</td>
<td>Very low primary bile acid concentrations</td>
<td>Low phospholipid concentrations</td>
</tr>
<tr>
<td><strong>Chromosomal locus</strong></td>
<td>18q21-22</td>
<td>2q24</td>
<td>7q21</td>
</tr>
<tr>
<td><strong>Gene/protein</strong></td>
<td>( ATP8B1/FIC1 )</td>
<td>( ABCB11/BSEP )</td>
<td>( ABCB4/MDR3 )</td>
</tr>
<tr>
<td><strong>Hepatocyte location</strong></td>
<td>Canalicular membrane</td>
<td>Canalicular membrane</td>
<td>Canalicular membrane</td>
</tr>
<tr>
<td><strong>Other main sites of expression</strong></td>
<td>Cholangiocytes, intestine, pancreas</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td><strong>Extrahepatic symptoms</strong></td>
<td>Diarrhea, pancreatitis, hearing loss, small stature</td>
<td>None</td>
<td>None</td>
</tr>
<tr>
<td><strong>Functional defect</strong></td>
<td>ATP-dependent aminophospholipid flippase</td>
<td>ATP-dependent bile acid transport in bile</td>
<td>ATP-dependent phosphatidylcholine flippase</td>
</tr>
</tbody>
</table>

GGT: Gamma-glutamyltransferase; ALT: Alanine aminotransferase; AFP: Alpha-fetoprotein; BSEP: Bile salt export pump; FIC1: Familial intrahepatic cholestasis 1; MDR3: Class III multidrug resistance
It is also likely that FIC1 disease represents a continuum comprising intermediate phenotypes between the benign phenotype BRIC1 and the severe phenotype PFIC1 [1,5]. So far, there is no clear explanation for the phenotypic differences between patients with BRIC1 and those with PFIC1. Analyses suggest that mutations identified in patients with PFIC1 would severely disrupt protein function, whereas protein function would be only partially impaired in patients with BRIC1 [5]. Genotype - phenotype associations would probably be complicated as a dramatic variability in phenotypic presentation has already been identified in patients with BRIC1 with a common mutation. In addition, many patients with FIC1 disease are compound heterozygous, which would also complicate the identification of genotype - phenotype correlations [5]. Heterozygous ATP8B1 mutations have been identified in some cases of transient neonatal cholestasis and in intrahepatic cholestasis of pregnancy (ICP), which may be referred to as “ICP type 1” (ICP1) [1,12].

**PFIC type 2**

PFIC2 (previously referred to as “Byler syndrome”) is caused by mutations in the ABCB11 gene (also designated BSEP) [1-13]. The ABCB11 gene encodes the ATP-dependent canalicular BSEP in human liver and is located on human chromosome 2. BSEP protein, expressed at the hepatocyte canalicular membrane, is the major exporter of primary bile acids against extreme concentration gradients. Mutations in this protein are responsible for the decreased biliary bile salt secretion described in affected patients, leading to decreased bile flow and accumulation of bile salts within the hepatocyte with ongoing severe hepatocellular damage. In addition, BSEP deficiency represents a phenotypic continuum between BRIC2 and PFIC2 [14]. So far, no clear genotype - phenotype correlation has been identified among PFIC2 patients, but most children with BSEP mutations, regardless of mutation type, have no canalicular BSEP protein expression [13]. Severe phenotypes are often associated with mutations leading to premature protein truncation or failure of protein production. Insertion, deletion, nonsense and splicing mutations result in damaging effects, with patients exhibiting little or no detectable BSEP at the hepatocyte canalculus. Missense mutations are also common defects that can either affect protein processing and trafficking or disrupt functional domains and protein structure [13,15]. Thus, detectable BSEP expression does not preclude functional BSEP deficiency. Some mutations have been functionally characterized and confirm the defect in bile acid secretion [16].

In milder disease such as BRIC2, missense mutations predominate over those leading to failure of protein production, and the mutations are seen in less conserved regions than in the nucleotide-binding folds (NBFs), which contain the Walker A/B motifs and the ATP-binding cassette (ABC) signature. Cholelithiasis has also been reported in BRIC2 patients [14, and personal data]. In addition, heterozygous ABCB11 mutations have been identified in cases of ICP2 [17], drug-induced cholestasis [18] and transient neonatal cholestasis [1].

**PFIC type 3**

PFIC3 is caused by a genetic defect in the ABCB4 gene (also designated MDR3) located on chromosome 7. Class III multidrug resistance (MDR3) of P-glycoprotein (P-gp), a phospholipid translocator involved in biliary phospholipid (phosphatidylcholine) excretion (flippase), is predominantly, if not exclusively, expressed in the canalicular membrane of hepatocytes [4]. Cholestasis results from the toxicity of bile in which detergent bile salts are not inactivated by phospholipids, leading to bile canalulci and biliary epithelium injuries. The mechanism of liver damage in PFIC3 patients is most likely related to the absence of biliary phospholipids. Injury to bile canalulci and biliary epithelium is probably due to continuous exposure to hydrophobic bile salts, the detergent effects of which are no longer being countered by phospholipids, thereby leading to cholangitis. In addition, the stability of mixed micelles in bile is determined by a three-phase system in which the proper proportion of bile salts and phospholipids is necessary for maintaining solubility of cholesterol. The absence of phospholipids in bile would be expected to destabilize micelles and promote bile lithogenicity through crystallization of cholesterol, which might favor small bile duct obstruction. These cholangiopathy mechanisms fit well with histological findings such as ductular proliferation. PFIC3 represents an important example of canalicular transport defect leading to the development of cholangiopathy.

The phenotypic spectrum of PFIC3 ranges from neonatal cholestasis to cirrhosis in young adults [4,19]. In our experience, ABCB4 sequence analysis in PFIC3 patients has revealed a large number of different ABCB4 mutations characterized on both alleles in most cases. In one-third of cases, mutations gave rise to a truncated protein. When tested, no MDR3 P-gp could be detected by liver immunostaining in these patients. The lack of MDR3 protein could be explained in two ways. The truncated protein may be broken down rapidly after synthesis, giving rise to extremely low steady-state levels of the protein. However, the premature stop codon is more likely to lead to instability and decay of the ABCB4 mRNA. This latter explanation is supported by the near absence of ABCB4 mRNA on northern blotting of liver samples from multiple patients. The remaining two-thirds of patients had missense mutations, some of which were found in the highly conserved amino-acid sequences of the Walker A/B motifs involved in ATP-binding. Such amino-acid changes in the Walker A or B motif are generally not compatible with ATPase activity and transport processes. Alternatively, missense mutations might result in intracellular misprocessing of MDR3, as shown with other ABC transporters [20]. Indeed, such missense mutations were associated with a decreased level of MDR3 canalicular protein. Whatever the mechanism(s) involved, the low level of biliary phospholipids found in patients with missense mutations demonstrates the MDR3 functional defect [4]. Also, there is now strong evidence that, in addition to PFIC3, a predominantly heterozygous MDR3 defect may be involved in ICP3 [17], cholesterol gallstone disease [21], drug-induced cholestasis [22], and some cases of transient neonatal cholestasis, adult idiopathic cirrhosis and cholangiocarcinoma [19,23,24]. Furthermore, MDR3 deficiency could represent a clinical continuum, as a single patient may experience different phenotypes during the disease course, starting with cholesterol cholelithiasis, followed by ICP and ending with biliary cirrhosis [25].
Diagnosis and diagnostic methods

PFIC should be suspected in children with a clinical history of cholestasis of unknown origin after exclusion of other common causes of cholestasis (such as biliary atresia, Alagille syndrome, alpha-1-antitrypsin deficiency, cystic fibrosis, sclerosing cholangitis and extrahepatic bile duct obstruction) [1,2]. High serum bile acid concentrations exclude primary disorders of bile acid synthesis [26]. Patients with PFIC1 and PFIC2 have normal serum GGT activity, while patients with PFIC3 have high serum GGT activity. PFIC3 patients can also be distinguished from patients with PFIC types 1-2 in that they rarely present with cholestatic jaundice in the neonatal period, but do so later on in infancy, childhood or young adulthood instead. A combined clinical, biochemical, radiological and histological approach, together with liver immunostaining and biliary lipid analysis, allows classification of PFIC types and may lead to adequate molecular diagnosis.

Diagnostic methods

Radiological approach

Initial ultrasonography of the liver must be performed to exclude biliary tract disease. Typically, ultrasonography is normal, but may reveal a huge gallbladder. Sometimes, biliary stones may be identified. If performed, cholangiography may show a normal biliary tree, excluding sclerosing cholangitis, and allows bile to be collected for biliary lipid analysis [1-4].

Liver histology

This is important for diagnosis as it allows liver immunostaining to be performed. Typical features have been described above (see the clinical description section). When signs of biliary obstruction appear on liver biopsy, as may be seen in PFIC3, cholangiography should be performed to exclude sclerosing cholangitis [1-4].

Liver immunostaining

Commercially available MDR3 and BSEP antibodies allow liver immunostaining to be performed [1,3,4,13]. Absence of canalicular or mild immunostaining favors a gene defect. However, normal staining does not preclude a gene defect as a mutation may induce loss of function, but with normal synthesis and expression.

Biliary lipid analysis

This can be performed with gallbladder bile or with bile collected by duodenal aspiration (pure choledochal bile). In cases of gallbladder puncture, bile contamination by blood may prevent proper bile analysis. In cases of duodenal aspiration, bile dilution or bile contamination by alimentary phospholipids may also interfere with proper bile analysis. Biliary bile salt concentrations may be dramatically decreased (< 1mM) in PFIC2 patients and only mildly decreased in PFIC1 patients (3-8 mM) [3], whereas normal concentrations of biliary primary bile salts distinguish PFIC3 patients from those with PFIC1 or PFIC2 [1,4]. However, in PFIC3 patients, the biliary phospholipid level is dramatically decreased (1-15% of total biliary lipids; normal range: 19-24%). Biliary bile salt-to-phospholipid and cholesterol-to-phospholipid ratios are approximately fivefold higher than in wild-type bile. The residual percentage of biliary phospholipids appears to be directly related to the severity of the MDR3 mutation and, consequently, to the residual activity of MDR3 P-gp. Patients with "severe" mutations (nonsense, frameshift) have a low percentage of biliary phospholipids (<2%), while patients with missense mutations have a higher percentage of biliary phospholipids (±2%). In our experience, the threshold predictive of a positive response to ursodeoxycholic acid (UDCA) therapy in PFIC3 is represented by a percentage of biliary phospholipids of around 7%. Thus, a combination of abnormal MDR3 canalicular immunostaining and a low percentage of biliary phospholipids is highly suggestive of MDR3 deficiency.

Molecular analysis

Gene analysis usually involves DNA sequencing of the 27 coding exons and their splice junctions. In cases of silent or intronic mutations, a test in silico is helpful for predicting a potential splice defect that would still need to be confirmed by RNA analysis using real-time reverse transcription polymerase chain reaction (RT-PCR) and sequencing [27]. The use of a resequencing chip dedicated to gene cholestasis could also facilitate identification of a gene mutation [28].

Differential diagnosis

Several other autosomal-recessive PFIC-like diseases are recognized (Table 2). Liver diseases resembling PFIC with normal GGT activity have been identified as inborn errors in primary bile acid synthesis and represent distinct disorders [1,2,26], and other liver diseases resembling PFIC with normal serum GGT activity are also known. Familial Amish hypercholanemia represents a PFIC-like disorder not due to a primary defect of the transport system involved in bile formation, but caused by a tight-junction protein defect combined with a defect of primary bile acid conjugation [1,29]. Cholestasis is caused by impaired transport of unconjugated bile acids into bile and bile leakage into plasma through abnormal canalicular tight junctions, thereby increasing paracellular permeability. Another category of progressive cholestatic liver disease of childhood might be due to abnormal villin expression leading to loss of structural integrity of the canalicular microvilli, thus impairing biliary secretion system function [1]. A case of hypercholanemia due to mutations in the m-epoxide hydrolase gene (EPHX1) has also been reported [1]. Arthrogryposis - renal dysfunction - cholestasis syndrome is a complex disease resulting from mutations of VPS33B/VIPAR, which are involved in intracellular trafficking and targeting of apical proteins. These gene defects
PFIC may be effective in some patients, especially those with PFIC3 with missense mutations, who have less severe disease in comparison to children with a mutation leading to truncated protein [4]. Some patients with PFIC1 or PFIC2 may also benefit from surgical biliary diversion [33]. Nasobiliary drainage may help to select potential responders to biliary diversion [34]. So far, however, clear genotype–phenotype correlation data are lacking and still need to be defined to identify those PFIC1–2 patients who might benefit from UDCA or biliary diversion [3]. Preliminary data suggest that PFIC2 patients with p.D482G or p.E297G mutations may respond well to biliary diversion. Nevertheless, if these therapies fail, liver transplantation represents the only alternative [1]. In our experience, the extrahepatic features, such as diarrhea, liver steatosis and short stature, sometimes associated with PFIC1 do not improve or may be aggravated following successful biliary diversion or liver transplantation [1]. In fact, chronic diarrhea may become intractable when biliary bile salt secretion is restored after liver transplantation [1,11]. It is often associated with severe liver steatosis and/or steatohepatitis that can lead to cirrhosis over time and to indications for retransplantation. Liver steatosis and diarrhea may even recur after retransplantation (personal data). Diarrhea may be favorably managed by bile adsorptive resin treatment [1,11]. In the future, therapies such as

| Table 2 Other progressive familial intrahepatic cholestasis (PFIC)-like diseases |
|-------------------------------|-------------------|-----------------|
| **Disease**                  | **GGT**           | **Protein, gene (chromosome)** |
| Primary bile acid synthesis or conjugation defects |                      |                              |
| 3β-HSD defect                | N                 | 3β-HSD, HSD3B7 (16)           |
| Δ4-3oxo-R defect             | N                 | Δ4-3oxo-5β R, AKR1D1 (7)      |
| Oxysterol 7α-OHase defect    | N                 | oxysterol 7α-OHase, CYP7B1 (8) |
| BAAT defect                  | N                 | BAAT, BAAT (9)               |
| BACL defect                  | N                 | BACL, SLC27A5 (19)           |
| Trafficking and canalicular targeting defects |                      |                              |
| ARC syndrome                 | N                 | VPS33B, VPS33B (15)          |
| Familial Amish hypercholanemia | N             | VIPAR, VIPAR (14)            |
| Tight-junction defects       |                   |                              |
| NISCH*                       | E                 | Claudin-1, CLDN1 (3)         |
| Familial Amish hypercholanemia* | N             | ZO-2, TJP2 (9)               |
| Other defects                |                   |                              |
| Familial hypercholanemia     | N                 | EPHX1, EPHX1 (1)             |
| NAICC                        | E                 | Cirhin, CIRH1A (16)          |
| PCC                          | E                 | Villin, ? (2)                |
| Aagenaes syndrome            | E                 | ?, (15q26.1)                 |

GGT: Serum gamma-glutamyltransferase activity; N: Normal; E: Elevated; 3β-HSD: 3β-hydroxy-Δ5-7-steroid oxydoreductase; Δ4-3oxo-R:Δ4-3-oxosteroid 5β-reductase; Oxysterol 7α-OHase: Oxysterol 7α-hydroxylase; BAAT: Bile acid-CoA:amino-acid N-acyltransferase; BACL: Bile acid-CoA ligase; ARC: Arthrogryposis – renal dysfunction – cholestasis; EPHX1: Epoxyde hydroxylase 1; NISCH: Neonatal ichthyosis sclerosing cholangitis; NAICC: North American Indian childhood cirrhosis; PCC: progressive cholestatic liver disease of childhood; *Claudin-1 defect is present in hepatocytes and cholangiocytes; ZO-2 defect is associated with a bile acid conjugation defect (BAAT defect)

result in a loss of apical protein expression in the liver and kidneys [30]. For the liver, it represents another PFIC-like disorder with normal GGT activity. In cases of high serum GGT activity, sclerosing cholangitis must be excluded [31]. North American Indian childhood cirrhosis due to a defect in the cirhin gene is characterized by high GGT activity, as is Aagenaes syndrome, a lymphedema cholestasis syndrome of unknown cause [1].

Genetic counseling and antenatal diagnosis

Genotyping should be used to confirm the diagnosis of PFIC in affected children. Heterozygosity of parents for the defects found in affected patients confirms the recessive inheritance of the disease. This awareness has already allowed prenatal diagnosis. However, antenatal diagnosis of PFIC requires clinical and biochemical expertise [23].

Specific management, including treatment

Therapy with UDCA should be considered in the initial therapeutic management of children with all types of PFIC [3,4,32]. UDCA therapy may be effective in some patients, especially those with PFIC3 with missense mutations, who have less severe disease in comparison to children with a mutation leading to truncated protein [4]. Some patients with PFIC1 or PFIC2 may also benefit from surgical biliary diversion [33]. Nasobiliary drainage may help to select potential responders to biliary diversion [34]. So far, however, clear genotype–phenotype correlation data are lacking and still need to be defined to identify those PFIC types 1–2 patients who might benefit from UDCA or biliary diversion [3]. Preliminary data suggest that PFIC2 patients with p.D482G or p.E297G mutations may respond well to biliary diversion. Nevertheless, if these therapies fail, liver transplantation represents the only alternative [1]. In our experience, the extrahepatic features, such as diarrhea, liver steatosis and short stature, sometimes associated with PFIC1 do not improve or may be aggravated following successful biliary diversion or liver transplantation [1]. In fact, chronic diarrhea may become intractable when biliary bile salt secretion is restored after liver transplantation [1,11]. It is often associated with severe liver steatosis and/or steatohepatitis that can lead to cirrhosis over time and to indications for retransplantation. Liver steatosis and diarrhea may even recur after retransplantation (personal data). Diarrhea may be favorably managed by bile adsorptive resin treatment [1,11]. In the future, therapies such as

- [30]
- [31]
- [32]
Progressive familial intrahepatic cholestasis (PFIC) patients who became pregnant [22,42]. Patients with BSEP deficiency, especially those with biallelic truncating mutations, are at considerable risk of hepatobiliary malignancy (hepatocellular carcinoma or cholangiocarcinoma) [3,13,41,43]. These findings justify close monitoring of hepatocellular carcinoma (at the very least with serum alpha-fetoprotein doses every 6 months and liver ultrasonography every year) in PFIC2 patients from the first year of life. Liver tumor monitoring is also justified in PFIC3 [4,24].

Questions to be answered in the future

In rare PFIC cases (< 10%), only one mutated allele or no mutation is identified [3,4]. This may be explained by mutations that map to the regulatory sequences of genes. A gene involved in the transcription of PFIC genes (such as FXR) or in trafficking/targeting of canalicular transporters (such as CDC50, Rab11a and myosin Vb) may also be involved [30,44–47]. In addition, it is possible that other, unidentified genes involved in bile formation are responsible for the PFIC types 1–3 phenotypes. Furthermore, it may be

Prognosis

In addition to the natural history and complications of all three types of PFIC (portal hypertension, liver failure, cirrhosis, hepatocellular carcinoma, extrahepatic manifestations), children with PFIC types 1-3 are also theoretically at risk of developing biliary stones, drug-induced cholestasis and ICP further along in the disease course. Girls receiving UDCA therapy who reach adulthood with their native liver should not stop UDCA during pregnancy because of the risk of developing severe ICP, as seen in a previously reported patient...
hypothesized that combined heterozygous mutations for two genes (say MDR3 and BSEP) may lead to a PFIC-like phenotype [48]. Another interesting possibility is that, in a heterozygous state, the mutated protein may have a dominant negative effect on protein expression/function [49]. Modifier genes and environmental influences could also play a role in the expression of PFIC.

After liver transplantation, the possible recurrence of PFIC on liver graft due to alloimmunization of the recipient against the BSEP, MDR3 or FIC1 proteins of liver donors remains a theoretical matter of debate. It is hypothesized that PFIC patients with severe mutations leading to a lack of gene product would be immunologically naive for FIC1, BSEP or MDR3 gene products. In this context, alloimmunization could arise following liver transplantation. To our knowledge, this post-transplant complication has been reported so far only in PFIC2 [50], manifesting as an unexplained severe bout of pure hepatocellular cholestasis resembling PFIC2. As yet, the incidence and natural history of this alloimmunization process is not well known.

In addition, in cases of parental living donor transplantation, it could be expected that the heterozygous status of the liver graft may lead to a predisposition for developing lithiasis or cholestasis favored by immunosuppressive drugs [18] that could interfere with canalicular protein function, as seen in a PFIC2 patient [1]. Nevertheless, this complication is likely to be extremely rare, as it has not been reported in a large case series of living donor transplantation for PFIC [51].

Competing interests

The author declares that he has no competing interests.

References


